

CLEANING LABORATORY EVALUATION SUMMARY

SCL #: 2024
 DateRun: 01/01/1970
 Experimenters: Aditi Patel, Namrata Chauhan, Serena Burkinshaw
 ClientType: Lab
 ProjectNumber: Project #12
 Substrates: Stainless Steel
 PartType: Coupon
 Contaminants: MS2 Bacteriophage
 Cleaning Methods: Pour Plate
 Analytical Methods: Organism count
 Purpose: To evaluate the efficacy of cleaners with inactivating MS2 on a hard surface without agitation

Experimental Procedure: Pour Plate Method - MS2 Bacteriophage
 Six hours prior to the run, E.coli 15597 was subcultured into three milliliters of tryptic soy broth (TSB) screw-cap tubes and incubated at 37°C (98.6°F). 27 screw-cap tubes filled with 10ml of 0.5X tryptic soy agar (TSA) were autoclaved. The biosafety cabinet (BSC) was sprayed with 70% v/v isopropyl alcohol using a paper towel before spraying any items going into the BSC. Once autoclaving was complete, the TSA tubes were placed into a 45°C (113°F) D.I. water bath inside the biosafety cabinet (BSC). The four glass Petri dishes were marked using a black sharpie to designate the positive (P+), negative (N-), Test 1 (T1), and Test 2 (T2). Ten microliters of the organism were pipetted onto the P+, T1, and T2 stainless steel coupons and air-dried for 15 minutes. A motorized pipette with 10ml tips was used to pipet 15 ml of Dey-Engley (D/E) neutralizing broth into four separate 50ml conical tubes labeled P+, N-, T1, and T2. Once the MS2 bacteriophage dried on the coupons, the P+ coupon was placed into the conical tube. The N-, T1, and T2 were pipetted with 1000µl of the cleaning solution onto each coupon for 30 seconds before immediately placing them in the conical tube with an autoclaved forceps. The conical tubes were then placed on the shaker for 10 minutes. During this time, using the 1000ml pipette, 900ml of 1x phosphate-buffered saline (PBS) was pipetted into nine autoclaved dilution tubes, and serial dilutions were made for P+, T1, and T2 up to 10⁻⁴ using 100µl of the shaken D/E broth. Once the six-hour sub-time was complete, the E. coli 15597 subculture was removed from the incubator for use. For each variable (N-, P+, T1, and T2), 100µl of the stock and serial dilutions of MS2 bacteriophage, and 100µl of the E.coli 15597 subculture were combined into an empty dilution tube. A screwcap tube of 0.5X TSA was removed from the water bath, wiped with a paper towel to remove moisture, and poured into the dilution tube. The mixture was immediately poured into a sterile polystyrene petri dish; swirled to cover the entire plate surface, and then air-dried before covering. Dried Petri dishes were placed into a clean labeled zip lock bag that was partially closed and incubated at 37°C overnight. Plates were counted the following day based on the clear lysis zones in the bacterial lawn of growth (1 plate forming unit) to calculate log reduction and percent removal.

Results:	Product Name	Log Reduction	Percent Reduction
	Mr. Clean Antibacterial-Summer Citrus	0.4929	74.9205
	Mr. Clean Multi-Surface Cleaner - Meadows & Rain	1.9896	84.2780
	Mr. Clean Clean Freak Multipurpose Cleaner	0.6022	52.1422

Summary:	Substrates: Stainless Steel		Contaminants: MS2 Bacteriophage			
	Company Name:	Product Name:	Conc.:	Efficiency:	Effective:	Observations:
	Mr Clean	Mr. Clean Antibacterial Cleaner Summer Citrus	100		<input type="checkbox"/>	
	Mr Clean	Mr. Clean Multi-Surface Cleaner Meadows & Rain	100%	84.28	<input type="checkbox"/>	
	Mr Clean	Mr. Clean Clean Freak Multipurpose Cleaner	100%	52.14	<input type="checkbox"/>	

Conclusion: None of the Mr. Clean cleaners effectively eliminated MS2. Mr. Clean Multi-Surface cleaner in meadows and rain scent had the highest percent reduction at 84.2780%.